

# Mechanisms for macroscopic chirality in organisms

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## Preface: why this topic for Landau?

Biological physics ... necessarily “pathological”?

In my other work – frustrated spins, toy fermionson lattices, quasicrystals – I try to explicitly connect microscopic Hamiltonian to coarse-grained parameters, opposite to Landau’s spirit.

But this topic deduces **physical** consequences from a priori, **fundamental symmetry principles**.

# Outline

Need **physical** mechanism to get chirality from micro to macro scale.

1. Introduction:

- Examples

- Assumptions/facts

- Classifying mechanism: cell/collective level)

2. Review of vertebrate case: cilia-driven flow

3. LR asymmetric transport of chemicals?

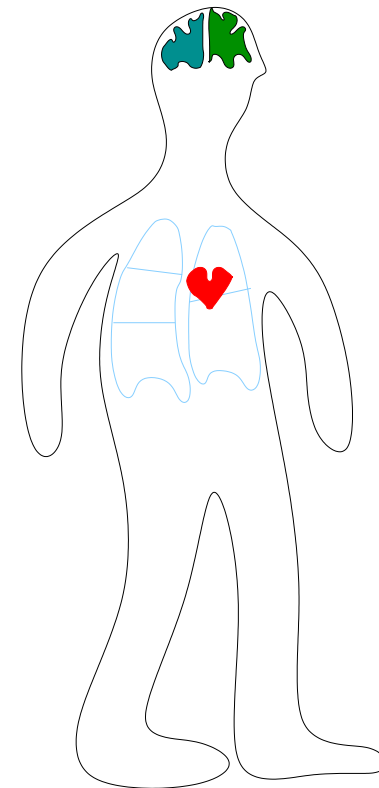
4. Twist in cell division

5. Conclusions

# 1. INTRODUCTION

## 1.1 Examples

- a) Vertebrates: heart, lungs, etc.  
(fraction reversed:  $\sim 10^{-4}$ )  
Model species: chick, Xenopus frog,  
zebrafish, mouse
- b) Human brain: left-hemisphere dominant /  
= “right-handed”  
(fraction reversed:  $\sim 10^{-1}$ )



c) *C. elegans* (nematode “worm”) and flies: twisting of gut (sorry, no image)

d) molluscs: shell twists right-handed  
Image: mating is awkward for R snail and (rare) L snail



No universal answer expected! (This is biology!).  
Different mechanisms (likely) apply to different cases.

## 1.2 Question and starting assumptions/fact:

An embryo develops 2 axes: **head/tail**=x, and **front/back**=z, by **spont. symmetry breaking**.

How to ensure 3rd axis (y) fulfills “right-hand rule” ( $\mathbf{y} = \mathbf{z} \times \mathbf{x}$ )?

**Symmetry**  $\Rightarrow$  **need all 3**: **x** and **z** axes, also (fibers’) **chirality**)

Take as given:

- 1 From genetic control + chirality of molecules  
(**not** shape of mother’s womb/egg)
- 2 L/R, too, is **spontaneous** symmetry breaking (usually);  
systematic bias amplified from **small** symm. breaking field.  
 $\Rightarrow$  a small L/R ( $O(10^{-2}?)$ ) effect will suffice
- 3 Seek the **earliest** L/R asymmetry  $\Rightarrow$  whatever makes the  
asymmetry, preceded it and was (**functionally**) symmetrical

## 1.3 Claim: it's not a “biological” mechanism

Usual “biological” mechanism:

transport/diffusion of signaling molecules + reactions

⊗ diffusion doesn't distinguish handedness

Requires a **physical** mechanism:

semi-macroscopic elements: long fibers

(yes! in each cell's cytoskeleton)

forces and torques (molecular motors that run on the fibers)

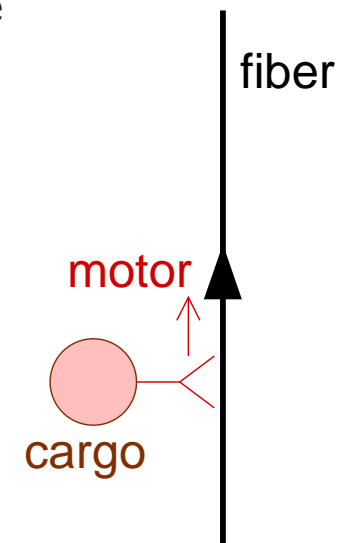
## 1.4 Review of the cytoskeleton

Cytoskeleton: Framework in each cell (of higher organisms), built from long, stiff, **directed**, **helical** macro-molecules (“fibers”)

- a) microtubules → motors dynein or kinesin
- b) actin → motor myosin

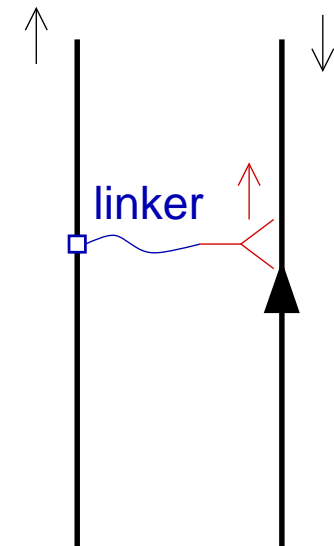
Each motor (literally) walks in a fixed sense (dissipating energy) carrying cargoes (chemicals in vesicles = little membrane bags)

Rarely (by thermal fluctuation) motor falls off its fiber;  
→ diffuses till it reattaches to some fiber





In place of a cargo, motor can be attached to another fiber, so one moves relative to other.  
(e.g. muscles).



There are many flavors within each kind of motor  
(used for specialized purposes)

## 1.5 Classifying mechanisms

(1) Two levels in any mechanism

- a) intracellular
- b) collective (intercellular): plan of the body's tissues

(2) Two styles in development (for different groups of animal):

a) **early: (many) invertebrates**

as cells first divide, each gets a determining label  
⇒ primarily cell-level mechanism suffices for L/R

b) **late: vertebrates**

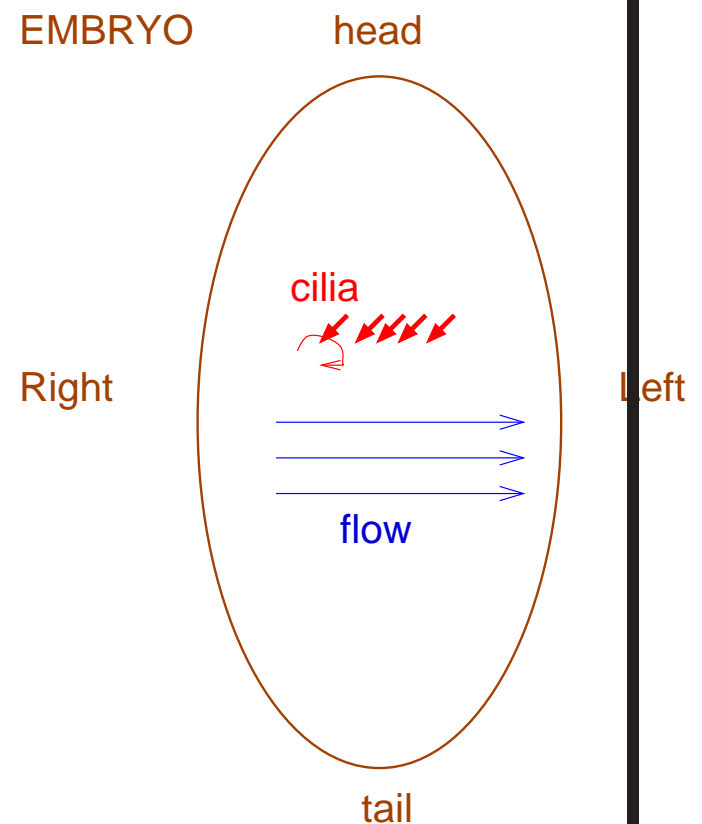
cells' fate isn't fixed early but pattern formation later  
⇒ need an inter-cellular mechanism

## 2. NODAL FLOW MECHANISM

Review: a late-stage mechanism in vertebrates

### 2.1 Cell-level story

Special cells on **front** of node (a key place on the embryo) have cilia:  
(moving tails that stick out from surface)  
tilted towards embryo's **tail** end.  
Cilia move **circularly** (CW looking down).  
Check: **all 3 ingredients** were mentioned.



Why node-cilia move circularly? Plausible from structure:

cilium has 9 pairs of microtubules that run its length  
each pair is linked by dynein molecules oriented clockwise  
(assembles from a microscopic template in the cell membrane)

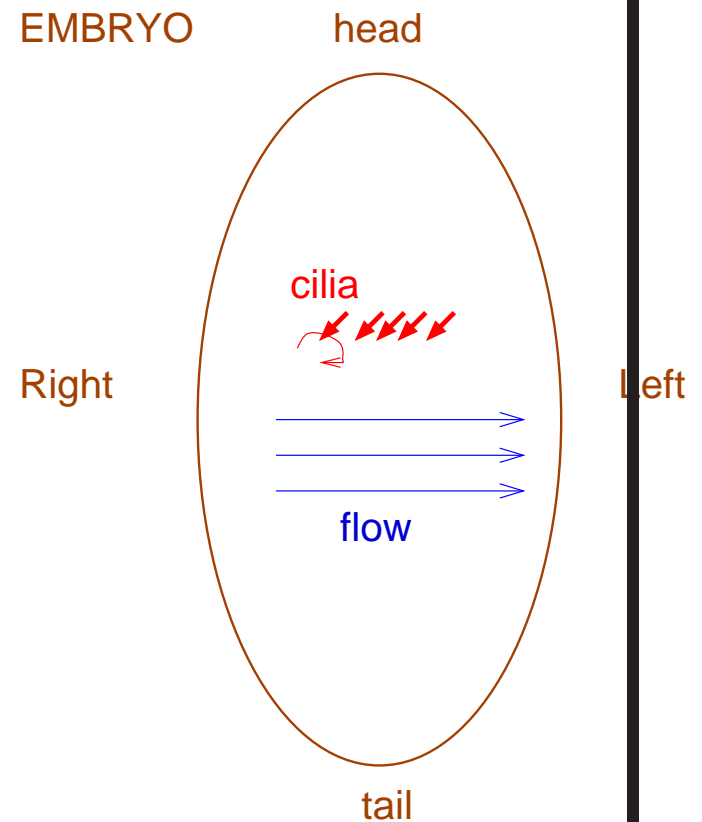
The interesting physics would be:

how this structure leads to circularly polarized motion  
(usual cilia move back and forth, not in circles)

Would need elastic theory + fluid dynamics.

## 2.2 Collective-level story

Sufficient to break symmetry:  
fluid flow gets driven **R to L**  
if a signaling chemical (still unidentified)  
is released, flow carries it to L side.  
Exp't: **force flow** the wrong way  
⇒ **reversed** embryos come out (in **mice**).



On the other hand, **earlier** L/R asymmetries are seen (in **frogs**)  
(Maybe early symmetry breaking has no functional importance?)

### 3. ASYMMETRIC TRANSPORT?

A “biological” mechanism uses **transport** of signaling molecules.

Can we have a **late-stage**, transport-based mechanism?

- Use: fiber molecules with motors (active transport works thus))
- Consider for simplicity a planar geometry (front and back sides)  
(indeed embryos are flattish at an intermediate stage)

Aim for: a “Hall effect” analog:

head-tail transport direction gets rotated by small angle.

**Cell level** is the nontrivial level for this mechanism.

## 3.1 Single cell level

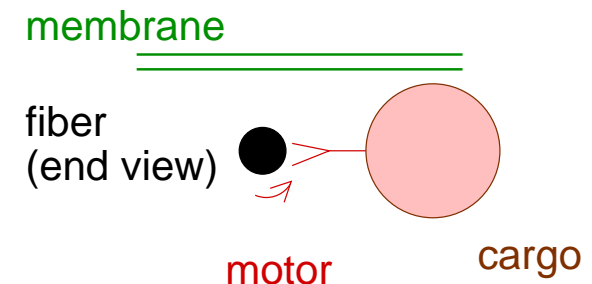
Assume (as common) array of fibers **just under** cell's membrane.  
(There's the **front/back asymmetry**)

Assume fibers oriented exactly along  $\pm z$ .  
(**not** (yet) the head/tail asymmm.)

**Active transport** depending on motor motion

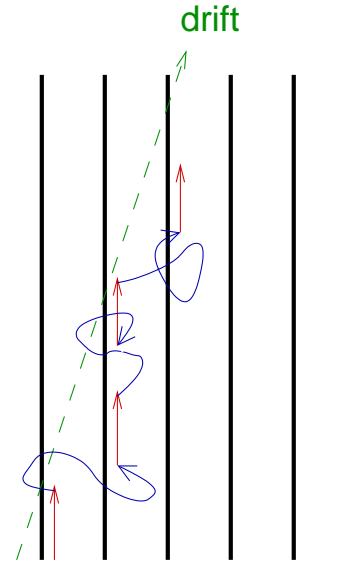
$\Rightarrow$  L/R **must** be due to the **generic helical component**  
of any motor's motion down any (microscopically) helical fiber.  
(There's the **chirality**.)

Say motor spirals **counter-clockwise**;  $\Rightarrow$   
pulls cargo till jammed against membrane.  
i.e. travels on **right** side of road



If cargo reattaches to a neighboring fiber, it's more likely to attach to the fiber on left side. Result: sideways drift: transport current at angle  $\theta$  from  $z$ , where

$$\theta \sim r_{\text{cargo}}/\ell \quad (1)$$



Here  $\ell \sim 1 \mu m$  is typical distance that motor goes before falling off, and  $r_{\text{cargo}} \sim 25 \text{ nm}$  is an effective distance of the cargo from the fiber axis (defined so  $r_{\text{cargo}}/d$  is probability of reattaching to the next fiber at a separation  $d$ )

So  $\theta \sim 2.5 \times 10^{-3}$ ? **Might** suffice as a biasing field...



## 3.2 Collective level

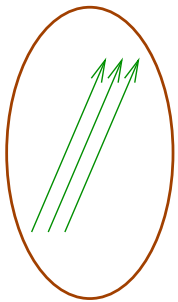
Now this array is only on the **front** side of the **front** layer of cells  
(Check: using front/back asymm. on organism level.)

Signal chemical is released from (say) tail end.  
(**Now** using **head/tail** asymmetry.)

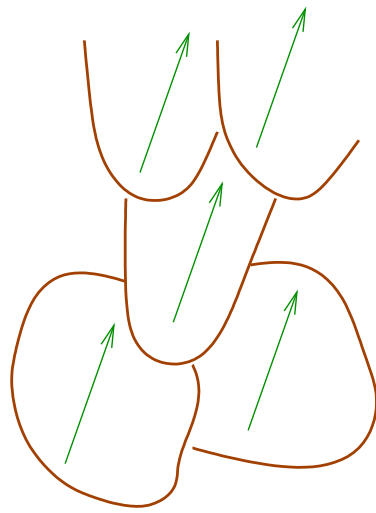
Then we argued signaling chemical has a **sideways bias**  
relative to the array in each cell

⇒ **macroscopic** transport has a similar bias

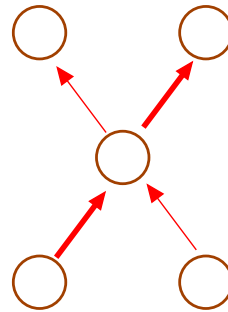
one cell



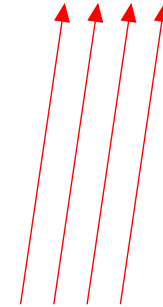
cells



discretized  
cells



coarse grained



**Summary:** it's not easy to generate late-stage asymmetry with a transport mechanism!

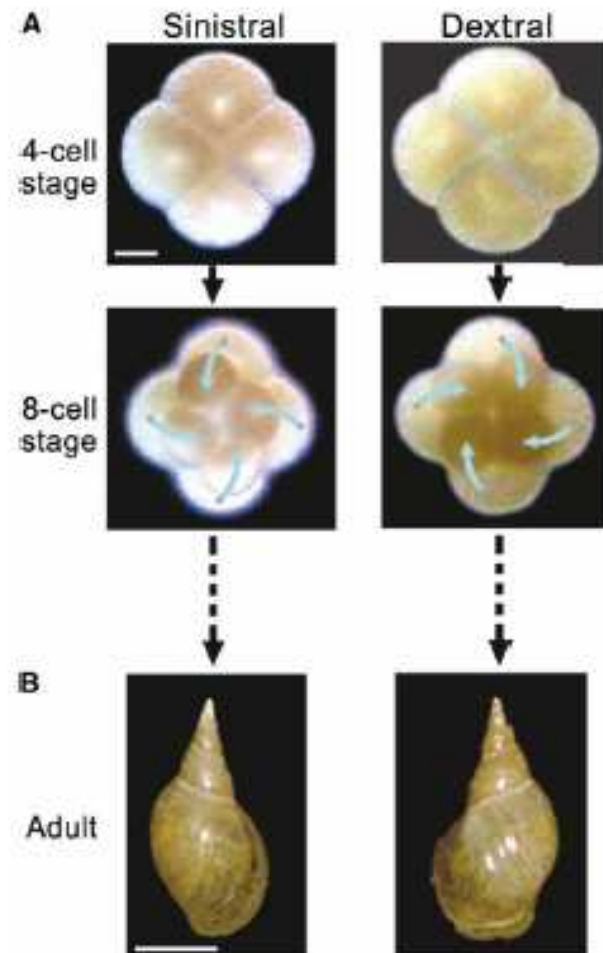
# 4. CELL DIVISION

Early-stage, cell-level mechanism.

## 4.1 Experimental facts:

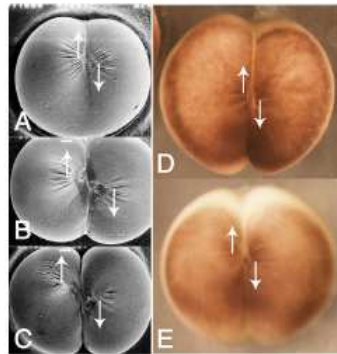
For Molluscs (snails).

- 1) 1st 4 cells in a square  
2nd 4 cells form on top with twist  
(normally in the sense shown).
- 2) Twist depends on **actin**  
but **not** on microtubules,  
of the 2 kinds of fiber.  
(?! cell division is driven by  
an array of **microtubules**)

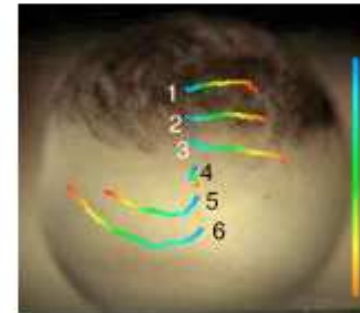


3) in frog eggs, same twist is observed and is due to actin; the parallel actin molecules shear past each other always in a clockwise sense; the motor myosin I-D is responsible

Actin forms a band pointing around the cell, in the plane where the split will occur;



Time lapse shows fluorescently labeled points along several fibers: uniform shear.



## 4.2 Model: asymmetric detachment

Start with array of parallel/antiparallel actin fibers.

Actin array's shear **must** be driven by the actin motor, myosin.

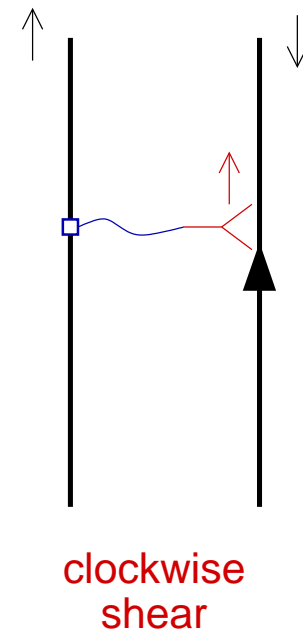
Each myosin walks on one actin fiber; it **must** be connected and bonded (directly or indirectly) to another fiber, in order to make any shear. I'll assume these connections are permanent bonds.

To drive clockwise (CW) shear, dominant bridging bonds **must** be placed as shown:  
(connect to R or to L, according to polarity of walked-on fiber).

[both ideas as in previous mechanism].

(1) How to get directed? **Must** depend on **helicity** of fiber, namely:  
myosin walks on actin with left-handed screw.

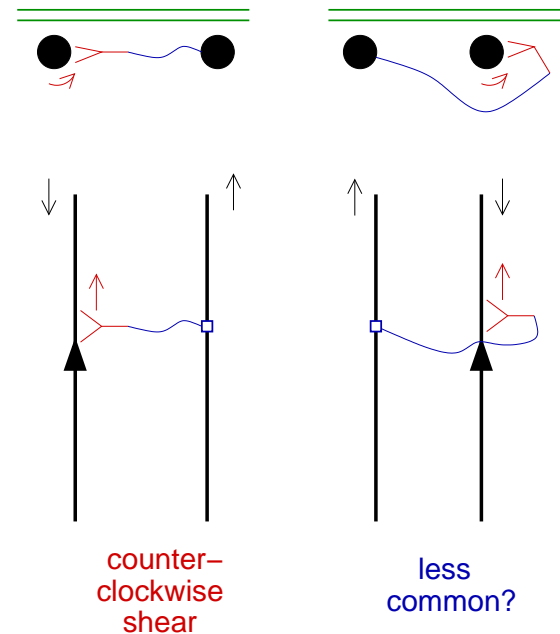
(2) Also, **must** depend on **front/back** difference; (probably)  
means interaction with **membrane**



Consider myosin motor bonded to fiber 1 and walking on fiber 2.  
 Motor spirals around fiber 2  
 till it comes against membrane  
 When motor is on **far** side of fiber 2,  
 linker strained  $\Rightarrow$  myosin falls off easier  
 $\Rightarrow$  near-side case dominates.

(Quantitative? depends on elasticity.)

Con: Predicts CCW! (wrong sense of shear)



## 5. CONCLUSIONS

Symmetry is key!

**Any** of the mechanisms to produce L/R asymmetry must explicitly use 3 ingredients:

**front/back** asymmetry, **head/tail** asymmetry, and **helicity** coming from the microscopic chirality of molecules.

Helicity generally comes by **screw mechanism**:

convert translation of **molecular motor** along **long fiber**

→ rotation around it

Universality of mechanism ?

Physics modeling allows less trivial universality:

may have same mechanism using different motors and fibers (myosin on actin → dynein on microtubules).

Biologists would tend to treat these as totally unrelated.



## Future experiments: suggestions

Study how motors carry cargo near a membrane

Study mutant forms of motor known to affect L/R:  
do they have changed screw motions around the fiber?

## (Near) future theory

Quantitative estimates for the various mechanisms  
(to test plausibility).

L/R in **plants** (no time in this talk!).

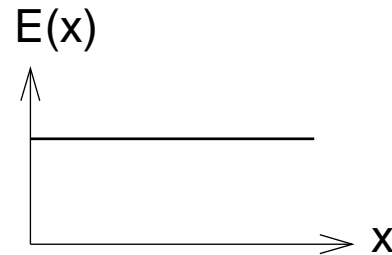
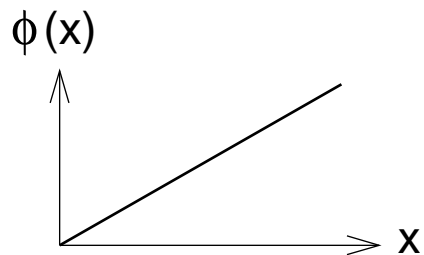
L/R in cell locomotion (nerve cells migrate far).

## [skip slide] 1.5(3)

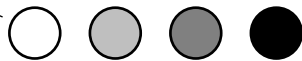
note too (3) Two ways to represent L and R

“positional information”: a polarization: a local vector

chemical conc.  $\phi(x, y, z) \propto x$ .  $\vec{E}(\mathbf{r})$ .



cells



Of course  $\vec{E}(\mathbf{r}) = \nabla\phi(\mathbf{r})$ . Nontrivial to convert:

- $\phi(\mathbf{r}) \rightarrow \vec{E}(\mathbf{r})$ : OK  
(Animal cells are big enough to sense conc. gradients between one side and the other)
- $\vec{E}(\mathbf{r}) \rightarrow \phi(\mathbf{r})$ : HARD  
(To get imbalance in conc.  $\phi(\mathbf{r})$  of a chemical,

it must be actively transported with a bias along  $\vec{E}(\mathbf{r})$

## [skip slide] 3.3 Alternate: array itself is twisted

How could an array get aligned with systematic twist  $\theta$ ? By collective behavior:

A fiber is nucleated at a random orientation at a certain rate per unit area; then it grows till it runs into some other fiber. that favors roughly parallel axes (in either sense).

Assume the fiber has a **right-hand** screw.

Say fiber 2 is growing and its end hits fiber 1. If it hits from the **right** [looking in the sense of the growing fiber 2], then fiber 2 gets carried **down** (by the screw thread of fiber 1); carried **up** if it hits from the **left**.

(here we used **helical** asymmetry!)

## [skip slide]

Imagine next that if fiber 2 gets carried **up** (hitting the membrane) it is more likely to **stop** growing; but if carried **down**, it's more likely to get carried under and beyond fiber 1, and **continue** growing. (Write difference as a small dimensionless factor  $f$ ) (here we used up/down asymmetry!)

Hence: new fibers at an orientation where they hit old fibers from the right grow longer; the new fibers' orientation will be rotated **counter-clockwise** by an angle of order  $f$ . (I'm glossing over possible logarithmic factors.) So, if the array started out aligned in the  $z$  direction, its later orientation is rotated by an angle  $\theta$  of order  $f$  times the number of generations of fiber growth since the start.

**[skip slide] 4.1**



## [skip slide] 4.3 Alternate Model: reshuffling of fibers

Cartoon: actin array is a mix of “new” fibers which are free, and “old” fibers which are crosslinked to each other (or otherwise attached to the cell), and **also** have more links bonded to them.

Myosin motor bonded to “old” fiber 1 walks along “new” fiber 2. When fiber 2 is on far side, elasticity of linker tends to pull fiber 2 around fiber 1 (which doesn’t move since it is attached to the cell). Thus, the only stable arrangement is when motor is on the near side, giving clockwise shear.

Pro: A more collective mechanism: with enough time, most fiber-fiber links are shearing in the same sense.

Con: Assumptions about formation of network.

Pro or con? Above version also predicts CCW shear; but may predict CW – depends on elastic assumptions?

## [skip slide] 4.4 Mutations can reverse the twist

Experimentally, in snails (or flies)

Note: reverse-twist behavior is **not** an exact mirror image: less strong?

Conjectures

- 1) mutant myosin motor twists around actin fiber in opposite sense, relative to its walking direction? (Note: some myosins walk in different senses.)
- 2) There are naturally two flavors of motor which twist in opposing senses. The mutation inactivates the dominant flavor, allowing the second flavor to win.
- 3) We had completely different mechanisms opposing each other. Mutation inactivates the myosin mechanism, allowing the



other] mechanism to control L/R.